1 Main Manuscript for

2

3	ZW sex chromosome differentiation in paleognathous birds is associated with
4	mitochondrial effective population size but not mitochondrial genome size or mutation rate
5	
6	Brooke Weinstein ¹ , Zongji Wang ² and Qi Zhou ^{2,3*} and Scott William Roy ^{1,4*}
7	¹ Department of Molecular and Cell Biology, University of California-Merced, Merced, CA
8	95343
9	² MOE Laboratory of Biosystems Homeostasis and Protection and Zhejiang Provincial Key
10	Laboratory for Cancer Molecular Cell Biology, Life Sciences Institute, Zhejiang University,
11	Hangzhou, China, 310058
12	³ Evolutionary & Organismal Biology Research Center, School of Medicine, Zhejiang
13	University, Hangzhou, Zhejiang, China, 310058
14	⁴ Department of Biology, San Francisco State University, San Francisco, CA 94117
15	* To whom correspondence should be addressed
16	Corresponding author:
17	Scott Roy
18	1600 Holloway Ave, San Francisco, CA 94132

19 415-602-4933

© The Author(s) 2025. Published by Oxford University Press on behalf of Society for Molecular Biology and Evolution. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

1 scottwroy@gmail.com

2 Keywords: evolution of complexity, purifying selection, Hill-Robertson Interference

3 Abstract

Eukaryotic genome size varies considerably, even among closely related species. The causes of 4 5 this variation are unclear, but weak selection against supposedly costly "extra" genomic 6 sequences has been central to the debate for over 50 years. The mutational hazard hypothesis, 7 which focuses on the increased mutation rate to null alleles in superfluous sequences, is 8 particularly influential, though challenging to test. This study examines the sex chromosomes 9 and mitochondrial genomes of 15 flightless or semi-flighted paleognathous bird species. In this clade, the non-recombining portion of the W chromosome has independently expanded stepwise 10 in multiple lineages. Given the shared maternal inheritance of the W chromosome and 11 mitochondria, theory predicts that mitochondrial effective population size (N_e) should decrease 12 due to increased Hill-Robertson Interference in lineages with expanded non-recombining W 13 14 regions. Our findings support the extent of the non-recombining W region with three indicators 15 of reduced selective efficiency: (1) the ratio of non-synonymous to synonymous nucleotide 16 changes in the mitochondrion, (2) the probability of radical amino acid changes, and (3) the 17 number of ancient, W-linked genes lost through evolution. Next, we tested whether reduced N_e 18 affects mitochondrial genome size, as predicted by weak selection against genome expansion. 19 We find no support for a relationship between mitochondrial genome size and expanded non-20 recombining W regions, nor with increased mitochondrial mutation rates (predicted to modulate 21 selective costs). These results highlight the utility of non-recombining regions and mitochondrial 22 genomes for studying genome evolution and challenge the general idea of a negative relation 23 between the efficacy of selection and genome size.

1 Significance

2 Explaining the striking variation in eukaryotic genome size and complexity has been a 3 long-standing challenge, primarily due to the need for well-controlled experiments. Using the 4 shared maternal inheritance of W chromosomes and mitochondrial genomes, we explore a novel 5 avenue for studying genome evolution. Our investigation of 15 paleognathous bird species, 6 which have experienced stepwise recombination suppression between the Z and W 7 chromosomes, introduces a powerful framework for unraveling genome structure evolution and confirms a compelling theoretical prediction: increased sex chromosome differentiation 8 9 correlates with decreased mitochondrial selective strength and efficiency. However, we find no 10 evidence of mitochondrial genome expansion under these conditions or with changes in mutation rate, calling into question that genome size and complexity are driven by differential selective 11 12 efficiency on nearly neutral superfluous genomic sequences.

13

14 Introduction

Nuclear and organellar genomes exhibit remarkable diversity in content and structure 15 across eukaryotes, characterized by significant variation in gene numbers, introns, gene copies, 16 17 and intergenic DNA (Smith and Keeling 2015). For over five decades, the hypothesis that 18 seemingly superfluous genomic elements persist in populations due to lack of weak or absent 19 selection (Ohta 1973, 1992; Doolittle 1978) has been central in debates about the origins of 20 genome size (GS) and complexity (Lynch 2007). Despite this long-standing discussion, the issue 21 remains unresolved, mainly due to technical complications, including challenges in estimating 22 key parameters such as the strength of selection. An alternative approach compares species with 23 varying effective population sizes (N_e) . N_e represents the theoretical number of individuals in an 24 ideal population experiencing genetic drift at a rate equivalent to the actual population,

2	that the relative influence of natural selection versus genetic drift varies with N_e , if genome
3	expansions are generally slightly harmful, they should be more likely to persist when N_e is small
4	(Lynch and Conery 2003; Lynch 2007).
5	The most extensively developed version of the conjecture that GS and complexity
6	differences reflect differences in the efficacy of selection is the mutational hazard hypothesis
7	(MHH; Lynch & Conery 2003). The MHH posits that alleles containing various types of
8	additional sequences (introns, extra gene copies, repetitive elements, etc.) will tend to have a
9	higher rate of mutation to null alleles, manifesting as a small non-zero cost relative to alleles
10	lacking the addition sequence. Thus, populations with small N_e will disproportionately
11	accumulate genomic insertions. In their original analysis, Lynch and Conery examined 43
12	genomes spanning prokaryotes, protists, fungi, plants, and animals. They estimated nucleotide
13	silent site diversity, π , predicted to equal $4N_e u$ or $2N_e u$ (for diploids and haploids, respectively,
14	where u represents the assumed constant per-nucleotide mutation rate), and found that $N_e u$
15	explained a significant portion of the observed variation in nuclear GS. However, objections
16	swiftly emerged on theoretical and methodological grounds.
17	Charlesworth and Barton (2004) highlighted challenges in accurately measuring N_e and
18	confounding effects with other aspects of organismal biology (such as development rate and
19	body size). Additionally, questions arose about whether microbes with large global N_e might
20	experience more substantial fitness effects from genome expansions than multicellular organisms
21	with smaller N_e (Batut et al. 2014). Furthermore, correcting for shared phylogenetic history
22	revealed that the perceived association between N_e and GS vanished (Whitney & Garland 2010).
23	Recent tests of the MHH have yielded elusive and contradictory results, primarily due to

accounting for ecological, demographic, and genomic complexities (Charlesworth 2009). Given

correlated evolutionary changes and ongoing debates over appropriate measures and definitions
 of *N_e* (Lefébure et al. 2017; Roddy et al. 2021).

3 Concerns about the challenges in measuring N_e directly in natural populations have been raised, so most studies rely on genetic or life history characteristics as proxies (James et al. 2017; 4 5 Waples et al. 2013). Beyond demographic factors, N_e at a genomic locus is influenced by the number of sites genetically linked to it. Deleterious mutations at these linked sites tend to remove 6 7 chromosomes from the population, effectively decreasing N_e (Charlesworth 2009). This is especially evident in differentiated non-recombining sex chromosomes (e.g., W and Y 8 9 chromosomes), where newly sex-linked non-recombining regions rapidly accumulate deleterious mutations and lose genes (Charlesworth and Charlesworth 2000). Consequently, comparing 10 genome evolution across related lineages with varying degrees of linkage is a promising 11 12 approach for testing the MHH. A compelling model system for investigating whether species with reduced N_e and lower 13 efficacy of selection evolve larger genomes comes from paleognathous bird sex chromosomes. 14 While most mammals share the same XY chromosome pair, nearly all birds possess a sex-linked 15 region within the ZW sex chromosomes. In most birds and all mammals, most of the sex 16 17 chromosome pair is differentiated. However, Paleognathae, the earliest diverging branch of 18 birds, represent an exception. Paleognathae, including flightless ratites and semi-flighted 19 tinamous, diverged from other birds over 110 million years ago (Jarvis et al. 2014). Intriguingly, 20 the Z and W chromosomes remain homomorphic in many paleognathous species, associated with 21 continued recombination in ZW females. Yet in multiple lineages, the non-recombining sex-22 linked portion of the chromosome has independently expanded due to stepwise, parallel 23 recombination suppression (Wang et al. 2022).

1	The power of this model system for studying GS evolution stems from the peculiar
2	inheritance of W chromosomes. Analogous to Y chromosomes in mammals, differentiated
3	regions of W chromosomes in birds are hemizygous and thus do not undergo recombination
4	(Charlesworth and Charlesworth 2000). Since the sex-specific (non-recombining) portion of the
5	W chromosome strictly follows maternal inheritance, it is expected to experience complete
6	genetic linkage with the entire mitochondrial genome (also maternally inherited). Rare or no
7	recombination occurs within either genomic element. Thus, reassortment between them is
8	expected to be absent or minimal, resulting in increased linkage predicted to amplify the effects
9	of background selection and hitchhiking, leading to increased Hill-Robertson interference (HRI)
10	and reduced N_e of the mitochondrial genome (Hill and Robertson 1966; Felsenstein 1974;
11	McVean and Charlesworth 2000; Santiago and Caballero 1995; Comeron et al. 2008;
12	Charlesworth and Jensen 2021). Some empirical support for reduced N_e during the expansion of
13	the sex-specific W region comes from observations of decreased nucleotide diversity in
14	mitochondrial genes from ZW species (Berlin et al. 2007). Parallel evolutionary ZW
15	differentiation across diverse paleognathous lineages, for which genomic data are available,
16	presents a rare opportunity to test the hypothesis that diminished selective efficiency leads to
17	larger genomes.
18	If the MHH holds, it should apply to all genomic features, including organelle GS.
19	However, Lynch et al. (2006) observed animals and land plants with similar N_e but dramatically
20	different mitochondrial GS and proposed an alternative hypothesis: organelle genome variation
21	results from differences in u rather than N_e , as previously suggested for nuclear genomes.
22	According to their argument, for mitochondria, the strength of selection, rather than its efficacy,

- 23 primarily drives GS and complexity evolution, with a lower *u* creating a more permissive
- 24 environment for accumulating additional hazardous DNA.

1	Similarly, as with Lynch & Conery's (2003) deployment of the MHH, support from
2	subsequent investigations into Lynch et al.'s (2006) proposal for organelle genomes have yielded
3	mixed support. Despite decades of research, the evolutionary forces governing GS variation
4	remain unclear. For a comprehensive review and discussion of competing hypotheses, refer to
5	Blommaert (2020) and Galtier (2024) for nuclear genomes and Smith (2016) for organelle
6	genome evolution. Before testing the two alternative predictions-GS expansion under either
7	decreased N_e or decreased u-we first confirmed the reduced N_e of the mitochondrial genome
8	under expanded ZW differentiation. We then examined whether increases in mitochondrial GS
9	variation are associated with enhanced drift in species with expanded regions of ZW
10	recombination suppression or decreased <i>u</i> .
11	
12	Results

Results 12

Differentiated W-linked genomic regions, inherited exclusively from mother to daughter 13 without recombination, mirror the inheritance pattern of mitochondrial genomes and are 14 completely genetically linked. The expansion of the non-recombining W-linked region is thus 15 expected to correlate with heightened HRI effects across both regions and the mitochondrial 16 17 genome, provided the expanded non-recombining segments harbor an increased complement of 18 functional sequences. To test this hypothesis, we obtained mitochondrial genomes and the 19 percentage of the sex chromosome pair that exhibits differentiation, along with a maximum-20 likelihood chronogram estimated from whole-genome non-coding sequences, for 15 species of 21 paleognathous birds, all previously generated by Wang et al. (2022). Among these species, 22 differentiated ZW regions vary widely in size, from 30% to 99% of the entire chromosome, with 23 a relative variation (the percentage ratio of the range to the average value) of 106%. Our initial 24 focus was on mitochondrial genes, so we created a concatenated alignment of the 13 proteincoding sequences across the 15 genomes. We found that the relative percent range in the proteincoding sequence (CDS) size is just 9.3% between species (ranging from 10.0 to 11.0 kilobases
(kb)). However, the size of intergenic regions varies considerably more, with a relative variation
of 153% (from 564 to 3815 base pairs). Overall, total mitochondrial genome size has a relative
range of 18.6% (from 15.8 to 18.9 kb; Figure 1).



6

Fig. 1. Paleognathae Phylogeny and Raw Mitochondrial Trait Data. Chronogram of the 15
paleognathous bird species analyzed. Heat maps depict the raw trait data. GS represents the total
mitochondrial genome size in kilobase pairs, while intergenic GS is the total GS minus the
coding region, tRNA, and rRNA genes. *u* is the estimated mutation rate per generation per base
pair. The red dot denotes the tinamou order/family. Branch lengths are in millions of years.

2 Diverse Evidence Suggests That ZW Sex Chromosome Differentiation Impacts the Effective 3 Population Size of W Chromosomes and Mitochondrial Genomes

We first sought to test whether the predicted relationship between the extent of ZW 4 5 differentiation and N_e holds. To assess the effect of ZW differentiation on mitochondrial gene evolution, we reconstructed branch-specific ratios of nonsynonymous changes (d_N; subject to 6 7 selection) to synonymous changes (d_s; presumed neutral) across the phylogenetic tree. Previous theoretical and empirical work shows that, under certain assumptions, the greater influence of 8 9 genetic drift under reduced N_e can lead to an overall increased fixation of deleterious 10 nonsynonymous variants. Consequently, d_N/d_S is expected to negatively correlate with N_e (Ohta 1992; Woolfit and Bromham 2005; Weyna and Romiguier 2021), resulting in a positive 11 relationship between the extent of recombination suppression on the W chromosome and d_N/d_S . 12 13 Consistent with this prediction, our ordinary least squares (OLS) regression analysis robustly confirms a positive association between the degree of ZW differentiation and d_N/d_S in 14 mitochondrial genes ($\beta = 0.035$, p-value = .012; Figure 2A). To ensure that the observed 15 16 variation in d_N/d_S is not solely due to differences in synonymous branch length, we tested for a 17 correlation between d_N/d_S and d_S . We found no evidence of such an association ($\beta = 0.0009$, pvalue = 0.962). Thus, as predicted, the size of the ZW differentiated region significantly 18 19 influences the mutation-normalized probability of fixation of nonsynonymous changes. 20 It is important to note that we did not apply phylogenetic correction in the model testing 21 d_N/d_S as affected by % ZW differentiation. The necessity and value of phylogenetic correction 22 arise from character states that apply to a given extant or ancestral taxon. These character states 23 exhibit phylogenetic inertia over time, remaining unchanged unless altered by evolutionary

1 processes (Felsenstein 1985). However, d_N/d_S is not a character state but a measure of 2 evolutionary change estimated from comparing a taxon and its direct ancestor. Consequently, the 3 concept of phylogenetic inertia does not apply to d_N/d_S (or the occurrence of radical amino acid 4 changes discussed below) in the same way. 5 Improper application of phylogenetic regression can lead to suboptimal statistical performance, especially when all the phylogenetic inertia is present in the predictor variable(s), 6 7 as noted by Rohlf (2006). Phylogenetic signal is generally assessed using Pagel's λ , which 8 ranges from zero to one, where values close to zero suggest the trait evolves independently of 9 phylogeny, and values near one indicate the trait evolves following Brownian motion along the branches. Within our data, all continuous variables exhibit moderate to strong phylogenetic 10 signals, represented by Pagel's λ , except for d_N/ds and the amount of intergenic DNA (left side 11 of Table I). Still, to be cautious, we followed Revell's (2010) recommendation to assess the 12 13 appropriateness of phylogenetic regression by testing whether model residuals contain a 14 phylogenetic signal, representing unexplained variation in the model associated with shared 15 evolutionary history. Again, while most regressions showed substantial signal, d_N/d_S, distributed 16 by % ZW, did not (right side of Table I). Revell (2010) also demonstrated that phylogenetic 17 correction is inappropriate and can be misleading under such circumstances. Therefore, we 18 corrected for phylogenetic inertia using phylogenetic generalized least squares (PGLS) for all 19 linear regressions except for the contribution of ZW differentiation to the d_N/d_S ratio. 20 21 Table I: Phylogenetic signal (λ) of the variables and estimated simultaneously on the 22 regression models' residuals, as Revel (2010) suggested. P-values are from hypothesis tests

for a significant phylogenetic signal against the null model ($\lambda = 0$).

Variable	λ	p-value	Regression model	λ	p-value
% ZW	1.00	0.0001	total GS ~ d_N/d_S	1.01	0.062
d _N /ds	0.00	1.00	$d_N/d_S \sim \% ZW$	-0.30	0.332
total GS	1.00	0.173	total GS ~ % ZW	1.00	0.031
intergenic GS	0.00	1.00	intergenic GS ~ % ZW	0.64	0.276
и	1.00	0.057	total GS ~ u	1.00	0.039
S0 genes lost	0.97	0.001	S0 genes lost ~ % ZW	0.94	0.001

In addition to an elevated accumulation of nonsynonymous mutations, a reduction in the 2 efficacy of selection is also predicted to correspond with increased occurrences of radical amino 3 acid changes, particularly those altering physicochemical properties like charge (Miyata et al. 4 1979), as these are more likely to affect protein function and be deleterious than conservative, 5 within-group, changes (Hanada et al. 2007). Indeed, binomial logistic regression revealed a 6 positive correlation between % ZW differentiation and the occurrence of radical amino acid 7 changes (Figure 2B). Specifically, we estimate that the log odds of an observed amino acid 8 change being radical increased by 0.47% with each one percent increase in ZW differentiation 9 10 (p-value = 0.044).

11 Additionally, we investigated whether ZW differentiation affects the N_e of the W 12 chromosome itself. To address this, we examined the retention and loss patterns of genes within 13 the ancestrally differentiated region of the chromosome (the oldest, "S0" stratum, as identified 14 by Xu and Zhou (2020)). The S0 stratum, shared by all birds, represents the most evolutionarily 15 ancient region of the W chromosome and is characterized by high sequence divergence and gene 16 loss, reflecting its long history of independent evolution. Expanded ZW differentiation predicts 17 increased HRI effects and thus decreased N_e for the S0 region. Consistent with increased HRI





1 Fig. 2. Confirmation of Reduced Selective Efficiency Due to Increased Recombination

2 **Suppression.** (A) A significant positive association exists between d_N/d_S and % ZW linkage. (B) 3 The predicted probability of an amino acid change being radical (estimated using binomial 4 logistic regression) increases with increasing % ZW linkage. Tick marks represent observations 5 of conservative (0; shown on the bottom axis) and radical (1; indicated on the top axis) amino 6 acid changes. β represents the logit coefficient, which is the change in the log odds of an amino acid change occurring being radical associated with every one percent increase in ZW linkage. 7 (C) A robust positive association is observed between the number of ancient W-linked S0 genes 8 9 lost and % ZW linkage. The blue line shows the regression coefficient (β) from ordinary least 10 squares regression (OLS), and the black line is from phylogenetic least squares regression 11 (PGLS) in 15 species of paleognathous birds.

12

13 Lack of Evidence for a Relationship Between Effective Population Size and Mitochondrial 14 Genome Size

If N_e significantly influences GS, the observed relationship between ZW differentiation 15 16 and N_e of the mitochondrion predicts a positive relationship between ZW differentiation and 17 mitochondrial GS. However, we found no correlation between % W chromosome differentiation 18 and mitochondrial GS, with neither PGLS ($\beta = 0.12$, p = 0.93) nor OLS regression ($\beta = 0.186$, p-value = 0.84) demonstrating any significant effect (Figure 3A), nor between mitochondrial GS 19 and d_N/d_S ($\beta = -6.16$, p-value = 0.70; Figure 3B). We also examined the relationship between 20 21 the amount of intergenic mitochondrial DNA and % W chromosome differentiation. We again 22 found no significant relationship with either PGLS ($\beta = 5.99$, p = 0.83) or OLS regression ($\beta =$

4 Lack of Evidence for a Relationship Between Mitochondrial Mutation Rate and

5 Mitochondrial Genome Size

Our dataset also allows us to test an additional proposed determinant of GS: u. According 6 7 to Lynch et al.'s (2006) model of organelle GS evolution, populations with higher u experience stronger selection against maladaptive increases in GS than those with lower u. To obtain 8 9 estimates of per-generation mitochondrial u, we first calculated the number of generations represented by each terminal branch length as the estimated branch length in years divided by the 10 extant species' estimated generation time. The reliability of fossil records in early-branching 11 birds has been well-scrutinized (Jarvis et al. 2014; Prum et al. 2015; Yonezawa et al. 2017), and 12 13 divergence time estimates are highly reliable and consistent, regardless of the taxon sampling and fossil calibrations used. We then computed per-generation mutation rates by dividing the 14 estimated d_s value by the estimated number of generations. It is important to note that while the 15 16 nuclear mutation rate influences branch lengths, the mitochondrial mutation rate does not affect 17 them. As these two parameters are governed by entirely different molecular machinery, we do 18 not expect a high degree of circularity when estimating mitochondrial u. We found no significant 19 relationship using PGLS ($\beta = 9.9e06$, p-value = 0.49) or OLS ($\beta = 3.1e06$, p = 0.83) (Figure 20 **3C**). Contrary to the predicted direction, the association was nonsignificantly positive, with 21 species with higher mutation rates tending to have larger genomes. These findings challenge the 22 predictions of the MHH and highlight the complex interplay of factors likely shaping genome 23 size in paleognathous birds.



1 Fig. 3. Correlations Between the d_N/d_S Ratio, Mitochondrial Genome Size, and Mutation 2 3 **Rate.** Blue lines represent the regression coefficient (β) from ordinary least squares regression (OLS), and black lines are from phylogenetic least squares regression (PGLS) in 15 species of 4 5 paleognathous birds. (A) No reliable correlation exists between d_N/d_S and mitochondrial genome 6 size (GS). (B) There is no reliable correlation between the amount of genetic linkage and 7 mitochondrial GS. (C) There is no significant relationship between the amount of intergenic 8 DNA and gene linkage. (D) No significant relationship is observed between mitochondrial GS 9 and the per-generation mutation rate per nucleotide site (u).

1 Discussion

2 Understanding the forces governing genome size has long captivated researchers. At the 3 forefront of this debate is the idea that nonessential genomic insertions are slightly deleterious, at 4 least under some circumstances, allowing them to fix and persist in some genomes while being 5 excluded from others. However, empirical tests of this hypothesis have been challenging due to 6 the multitude of potential factors influencing selection intensity against these elements 7 (Charlesworth and Barton 2004). Additionally, the technical complexities of quantitatively estimating these factors pose significant hurdles (Waples et al. 2013). Our study provides a rare, 8 9 relatively direct test of the slightly deleterious genome expansion hypothesis, relying on a theoretically and empirically supported decrease in selective efficiency resulting from increased 10 11 genetic linkage.

Our first major finding confirms the predicted association between increased ZW sex 12 chromosome differentiation and reduced N_e in genetically linked mitochondrial and sex 13 chromosomal genomic regions. While the predicted consequence of increased ZW differentiation 14 is a decrease in N_e due to greater HRI under increased genetic linkage, confidently inferring this 15 causality remains challenging as alternative explanations are plausible. Notably, a decrease in 16 17 population N_e could drive ZW differentiation and mitochondrial N_e . Some may interpret our 18 results in terms of HRI without invoking N_e . However, HRI and N_e are interconnected: N_e is 19 defined as nucleotide diversity (π) over u, and HRI reduces π , thereby implying changes in N_e. 20 Since HRI is one factor that influences N_e , differences in N_e due to varying levels of HRI are 21 pertinent to testing hypotheses about its overall effects. Our test relies on predicted directional 22 differences in the magnitude of N_e and the efficacy of selection rather than on specific estimates 23 of N_e .

1	Our second significant finding is that the GS of the mitochondrion is not associated with
2	the N_e or u of the mitochondrion. To clarify the relevance of both u and N_e in testing hypotheses
3	related to genome evolution, it is worth noting that the parameter $N_e \ge u$ is consistently
4	emphasized in population genetics, as it represents the combined influence of genetic drift and
5	mutation rate, which is central to understanding the balance between the introduction of new
6	mutations and the efficiency of selection in removing deleterious alleles. Fundamentally, the
7	MHH claims that "mutationally hazardous" DNA is more likely to accumulate in species with a
8	small N_e and low u when genetic drift is high than those with high N_e and u when drift is
9	minimal. While previous work often focuses on only one of the parameters – highlighting N_e
10	when discussing nuclear genomes and u when discussing mitochondrial genomes – the emphasis
11	on one variable over the other is a matter of choice by previous authors rather than a fundamental
12	issue with the underlying theory. Our failure to find a relationship between GS and N_e or u is
13	inconsistent with the prediction of the MHH hypothesis and the broader hypothesis that genomic
14	expansion incurs a fitness cost. Given the longstanding challenges in enacting controlled tests of
15	this hypothesis and the relatively straightforward nature of the natural experiment used here, our
16	results suggest a need for a more sustained effort to assess the predictive power of the MHH and
17	the idea that increased genome size and complexity are, by and large, slightly deleterious.
18	However, two potentially important objections may be raised to our approach. First,
19	mitochondrial genome size shows little variation across the studied taxa and thus may not
20	represent an ideal dataset. While it is true that total variation in genome size is moderate, there is
21	substantial variation in the overall contents and structure of the genome, from the highly
22	streamlined genome of Apteryx mantelli, where the core coding sequences account for the vast
23	majority of the genome (96%) to Struthio camelus, in which intergenic DNA makes up nearly a
24	quarter of the genome (20%). However, the fact that these taxa have relatively slight variation in

mitochondrial GS despite varying N_e itself rejects the MHH, given that it predicts variation under 1 2 these circumstances. It may be that bird mitochondrial genome size is governed by a cryptic 3 lineage-specific tendency towards smaller GS, driven by the energetic demands of flight (Wright et al. 2014), or, in the case of the semi-flighted or flightless birds studied here, the high 4 5 metabolic cost of running (Bundle et al. 1999). However, again, these contentions oppose the central claim of the MHH, namely that genome size variation is primarily explained by N_e or u 6 7 (Lynch and Conery 2006). Asserting that a hypothesis possesses explanatory power only under specific post hoc circumstances implies that its overall explanatory capacity remains limited at 8 9 best.

A second question concerns the extent of the change in N_e arising from the expanded W 10 chromosomal region. Estimating such values requires extensive knowledge of the distribution of 11 selective effects of newly arising nonsynonymous mutations in the mitochondrial genome. While 12 13 the increases in d_N/d_S may be seen as somewhat moderate (around 1.5 fold between short-W and long-W species), widespread degradation of W chromosome regions in birds and other lineages 14 (involving gene loss, increased d_N/d_S and the massive accumulation of transposable elements) 15 16 suggests substantial reductions in the efficiency of selection (Wang et al. 2021; Warmuth et al., 17 2022).

Our investigation represents a rare opportunity to explore a relatively well-controlled instance of the intricate relationship between genetic drift, mutation rate (u), and organelle genome size (GS). By focusing on a controlled case where variation in N_e arises from genetic changes in a single genomic region (rather than global demographic shifts), we confirm the expected changes in N_e through standard measures of selective efficiency. To transcend the specific framing of any single hypothesis, our study tested a broader prediction: if mitochondrial GS expansions are slightly deleterious, their fixation should increase as N_e or u decrease. 1 However, our failure to find the expected associations underscores the limitations of the

2 mutational hazard hypothesis (MHH) and other explanations positing weak or inefficient

3 selection on expanded genomes, highlighting the need for further controlled tests.

4

5 Materials and Methods

We estimated the mitogenome-wide d_N/d_S ratio using the FitMG94 workflow in HyPhy v. 6 2.5.36 (https://github.com/veg/hyphy-analyses/tree/master/ FitMG94); Pond and Muse 2005), 7 including credible intervals. The MG94 codon evolution model incorporates synonymous and 8 9 nonsynonymous nucleotide substitution rates as parameters, correcting for multiple hits at a codon and allowing ds to vary across branches (Muse and Gaut 1994). Additionally, the 10 FitMG94 workflow employs a corrected empirical estimator (CF3x4), which provides improved 11 estimates of several parameters from a standard model. This estimator accounts for individual 12 nucleotide frequencies at three codon positions and corrects for biases induced by stop codons 13 14 (Goldman and Yang 1994). To validate our HyPhy estimates, we compared them to d_N/d_S estimates obtained using 15

the free-ratio model in PAML (Yang 2007) and found they largely agreed. However, PAML 16 17 lacks a formal method for calculating credible intervals. Given the small sample size and 18 considerable uncertainty associated with both methods for the two short-branch sister taxa, we 19 opted to proceed with the Hyphy estimates. For the same reasons, we implemented Bayesian 20 linear and mixed phylogenetic models incorporating the uncertainty estimates for d_N/d_S . We 21 present the latter here since our Bayesian approaches yielded qualitatively similar results to 22 likelihood methods. Results and information on the Bayesian analyses are available in the 23 supplemental materials. Generation times used for estimating *u* were sourced from Wang et al. 24 (2021) unless otherwise specified.

1	To estimate ancestral states for % ZW differentiation, we used the fastAnc (fast
2	estimation of ML ancestral states) function in phytools (Revell 2012). We used aaML in PAML
3	without rate variation to infer ancestral mitogenome sequences. We then calculated radical and
4	conservative amino acid changes across all internal and external branches using RadAA (Seim et
5	al. 2019), which identifies pairwise amino acid changes in multiple sequence alignments and
6	categorizes residues into groups based on their charge, with cysteine forming its own group. The
7	lengths of tRNA genes were calculated using Arwen (Laslett and Canbäck 2008) and tRNAscan-
8	SE (version 2.0; Chan et al. 2021), while the ribosomal RNA genes (rrnL and rrnS) were
9	annotated using DeGeCI (version 1.1; Fiedler et al. 2024).
10	We computed individual variables' phylogenetic signal (λ) using the phylosig function in
11	phytools (Revell 2012). To validate the use of phylogenetic correction for linear regressions, we
12	followed Revell's (2010) instructions to simultaneously estimate Pagel's lambda (λ) with the
13	linear regression using the "corPagel" and gls function in nlme (Pinheiro et al. 2017). P-values
14	for the significance of the phylogenetic signal in the residuals were obtained using an ANOVA
15	test comparing our λ model with a model that has λ fixed at zero. Unless specified in the
16	supplemental materials, all statistical analyses were performed using R (v4.3.1).
17	
18	Acknowledgments

- BNW and SWR were supported by the National Science Foundation [grant number 1616878].
- 21 **Data Availability**
- 22 The primary data underlying this article are available on Github at
- 23 https://github.com/Brookesloci/Paleognath_ZW_GS.

1 Literature Cited

3	Batut, B., Knibbe,	C., Marais,	G., & Daubin,	V. 2014. Reductive	genome evolution at both ends
		, ,			0

- 4 of the bacterial population size spectrum. Nat. Rev. Microbiol., 12(12):841-850.
- 5 Berlin, S., Tomaras, D., & Charlesworth, B. 2007. Low mitochondrial variability in birds may
- 6 indicate Hill–Robertson effects on the W chromosome. Heredity, 99(4):389-396.
- 7 Blommaert, J. 2020. Genome size evolution: towards new model systems for old questions. Proc.
- 8 Royal Soc. B, 287(1933):20201441.
- 9 Bundle, M. W., Hoppeler, H., Vock, R., Tester, J. M., & Weyand, P. G. 1999. High metabolic
- 10 rates in running birds. Nature, 397(6714):31-32.
- 11 Chan, P. P., Lin, B. Y., Mak, A. J., & Lowe, T. M. 2021. tRNAscan-SE 2.0: improved detection
- 12 and functional classification of transfer RNA genes. Nucleic Acids Res., 49(16):9077-9096.
- 13 Charlesworth, B. 2009. Effective population size and patterns of molecular evolution and
- 14 variation. Nat. Rev. Genet., 10(3):195-205.
- 15 Charlesworth, B., & Charlesworth, D. 2000. The degeneration of Y chromosomes. Philos. Trans.
- 16 R. Soc. of Lond. B, Biol. Sci., 355(1403):1563-1572.
- 17 Charlesworth, B., & Barton, N. 2004. Genome size: does bigger mean worse?. Curr. Biol.,
- 18 14(6):R233-R235.
- Charlesworth, B., & Jensen, J. D. 2021. Effects of selection at linked sites on patterns of genetic
 variability. Annual review of ecology, evolution, and systematics, 52:177-197.
- 21 Comeron, J. M., Williford, A., & Kliman, R. M. 2008. The Hill–Robertson effect: evolutionary
- consequences of weak selection and linkage in finite populations. J. Hered., 100(1):19-31.
- 23 Doolittle, W. Ford. 1978. Genes in pieces: were they ever together? Nature, 272.5654:581-582.
- Felsenstein, J. 1974. The evolutionary advantage of recombination. J. Genet. 78(2):737-756.

- 1 Felsenstein, J. 1985. Phylogenies and the comparative method. Am. Nat., 125(1):1-15.
- 2 Fiedler, L., Bernt, M., & Middendorf, M. 2024. DeGeCI 1.1: a web platform for gene annotation
- 3 of mitochondrial genomes. Bioinform. adv., 4(1):vbae072.
- 4 Galtier, N. 2024. Half a Century of Controversy: The Neutralist/Selectionist Debate in Molecular
- 5 Evolution. Genome Biology and Evolution, 16(2):evae003.
- 6 Goldman, N., & Yang, Z. 1994. A codon-based model of nucleotide substitution for protein-
- 7 coding DNA sequences. Mol. Biol. Evol., 11(5):725-736.
- 8 Hanada, K., Shiu, S. H., & Li, W. H. 2007. The nonsynonymous/synonymous substitution rate
- 9 ratio versus the radical/conservative replacement rate ratio in the evolution of mammalian genes.
- 10 Mol. Biol. Evol., 24(10):2235-2241.
- 11 Hill, W. G., & Robertson, A. 1966. The effect of linkage on limits to artificial selection. Genet.
- 12 Res., 8(3):269-294.
- 13 James, J., Castellano, D., & Eyre-Walker, A. 2017. DNA sequence diversity and the efficiency
- 14 of natural selection in animal mitochondrial DNA. Hered. 118(1):88-95.
- 15 Jarvis, E. D. et al. 2014. Whole-genome analyses resolve early branches in the tree of life of
- 16 modern birds. Science, 346(6215):1320-1331.
- 17 Kimura, M. 1968. Evolutionary rate at the molecular level. Nature, 217(5129):624-626.
- 18 Kimura, M. 1983. The neutral theory of molecular evolution. Cambridge University Press.
- Laslett, D., & Canbäck, B. 2008. ARWEN: a program to detect tRNA genes in metazoan
 mitochondrial nucleotide sequences. Bioinformatics, 24(2):172-175.
- 21 Lefébure, T. et al. 2017. Less effective selection leads to larger genomes. Genome Res.,
- 22 27(6):1016-1028.
- 23 Lynch, M., & Conery, J. S. 2003. The origins of genome complexity. Science, 302(5649):1401-
- **24** 1404.

- 1 Lynch, M., Koskella, B., & Schaack, S. 2006. Mutation pressure and the evolution of organelle
- 2 genomic architecture. Science: 311(5768), 1727-1730.
- 3 Lynch, M. 2007. *The origins of genome architecture* (Vol. 98). Sunderland, MA: Sinauer
- 4 Associates.
- 5 McVean, A.T., & Charlesworth B. 2000. The effects of Hill-Robertson interference between
- 6 weakly selected mutations on patterns of molecular evolution and variation. Genetics 155.2:929-
- 7 944.
- 8 Miyata, T., Miyazawa, S., & Yasunaga, T. 1979. Two types of amino acid substitutions in
- 9 protein evolution. J. Mol. Evol. 12: 219-236.
- 10 Muse, S. V., and B. S. Gaut. 1994. A likelihood approach for comparing synonymous and
- 11 nonsynonymous nucleotide substitution rates with application to the chloroplast genome. Mol.
- 12 Biol. Evol. 11:715–724.
- 13 Ohta, T. 1973. Slightly deleterious mutant substitutions in evolution. Nature 246.5428:96-98.
- Ohta, T. 1992. The nearly neutral theory of molecular evolution. Annu. Rev. Ecol. Syst., p. 263286.
- Pinheiro, J. et al. 2017. Package 'nlme'. Linear and nonlinear mixed effects models, version,
 3(1).
- Pond, S. L. K., & Muse, S. V. 2005. HyPhy: hypothesis testing using phylogenies. In: *Statistical methods in molecular evolution*. Springer, New York, NY. p. 125-18.
- 20 Prum, R. O., Berv, J. S., Dornburg, A., Field, D. J., Townsend, J. P., Lemmon, E. M., &
- 21 Lemmon, A. R. 2015. A comprehensive phylogeny of birds (Aves) using targeted next-
- 22 generation DNA sequencing. Nature, 526(7574):569-573.
- 23 Revell, L. J. 2010. Phylogenetic signal and linear regression on species data. Methods Ecol.
- Evol., 1(4):319-329.

- 1 Revell, L. J. 2012. phytools: an R package for phylogenetic comparative biology (and other
- 2 things). Methods Ecol. Evol., 2:217-223.
- 3 Roddy, A. B., Alvarez-Ponce, D., & Roy, S. W. 2021. Mammals with small populations do not
- 4 exhibit larger genomes. Mol. Biol. Evol., 38(9):3737-3741.
- 5 Rohlf, F. J. 2006. A comment on phylogenetic correction. Evol. 60(7):1509-1515.
- 6 Santiago, E., & Caballero, A. 1995. Effective size of populations under selection. Genetics.
- 7 *139*(2):1013-1030.
- 8 Seim, I., Baker A., and Chopin L.K. 2019. RadAA: A Command-line Tool for Identification of
- 9 Radical Amino Acid Changes in Multiple Sequence Alignments. Mol. Inform., 38.1-2:1800057.
- 10 Smith, D. R. 2016. The mutational hazard hypothesis of organelle genome evolution: 10 years
- 11 on. Mol. Ecol., 25(16):3769-3775.
- 12 Smith, D. R., & Keeling, P. J. 2015. Mitochondrial and plastid genome architecture: reoccurring
- 13 themes, but significant differences at the extremes. Proc. Natl. Acad. Sci., 112(33):10177-10184.
- 14 Wang, Z. J., Chen, G. J., Zhang, G. J., & Zhou, Q. 2021. Dynamic evolution of transposable
- elements, demographic history, and gene content of paleognathous birds. Zool. Res., 42(1):51.
- 16 Wang, Z. et al. 2022. Phylogeny and sex chromosome evolution of Palaeognathae. J. Genet.
- 17 Genomics, 49(2):109-119.
- Waples, R. S., Luikart, G., Faulkner, J. R., & Tallmon, D. A. 2013. Simple life-history traits
 explain key effective population size ratios across diverse taxa. Proc. Royal Soc. B: Biol. Sci.,
 280:20131339.
- 21 Warmuth, V. M., Weissensteiner, M. H., & Wolf, J. B. 2022. Accumulation and ineffective
- silencing of transposable elements on an avian W Chromosome. Genome Res., 32(4):671-681.
- 23 Weyna, A., & Romiguier, J. 2021. Relaxation of purifying selection suggests low effective
- 24 population size in eusocial Hymenoptera and solitary pollinating bees. Peer Community J., 1.

- 1 Whitney, K. D., & Garland Jr, T. 2010. Did genetic drift drive increases in genome complexity?
- 2 PLoS Genet., 6(8):e1001080.
- 3 Woolfit, M., & Bromham, L. 2005. Population size and molecular evolution on islands. Proc.
- 4 Royal Soc. B: Biol. Sci., 272(1578):2277-2282.
- 5 Wright, N. A., Gregory, T. R., & Witt, C. C. 2014. Metabolic 'engines' of flight drive genome
- 6 size reduction in birds. Proc. Royal Soc. B: Biol. Sci., 281(1779):20132780.
- 7 Xu, L., & Zhou, Q. 2020. The female-specific W chromosomes of birds have conserved gene
- 8 contents but are not feminized. Genes, 11(10):1126.
- 9 Yang, Z. 2007. PAML 4: phylogenetic analysis by maximum likelihood. Mol. Biol. Evol., 24.8:
- 10 1586-1591.
- 11 Yonezawa, T. et al. 2017. Phylogenomics and morphology of extinct paleognaths reveal the
- 12 origin and evolution of the ratites. Curr. Biol, 27(1):68-77.